



## **AACE Italian Chapter Course 2**

*Official language: English*

### **Mineral metabolism disorders and osteoporosis: diagnosis and treatment**

#### **SESSION 1**

##### **Mineral metabolism disorders**

1. Hypercalcemia of malignancy:  
pathogenesis and management
2. Disorders of phosphate  
homeostasis
3. Paget's disease of bone
4. Metabolism of kidney stone  
disease
5. The bone-muscle-fat cross-talk

## **17° Congresso Nazionale AME**

Joint Meeting with AACE Italian Chapter

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# DISCLOSURE

▪

*I have no actual or potential conflict of interest  
in relation to this presentation.*



- ACKNOWLEDGEMENTS
- FAT, BONE AND MUSCLE CORRELATION: EVIDENCE BASED OBSERVATIONS
- FAT, BONE AND MUSCLE CORRELATION: FROM BASIC OBSERVATIONS TO POTENTIAL MECHANISMS OF INTERACTION
- TAKE HOME MESSAGES



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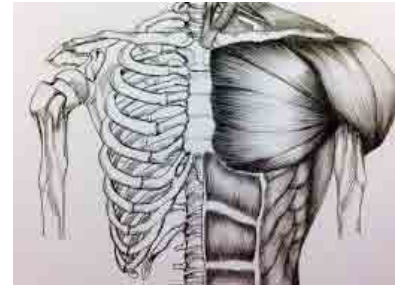
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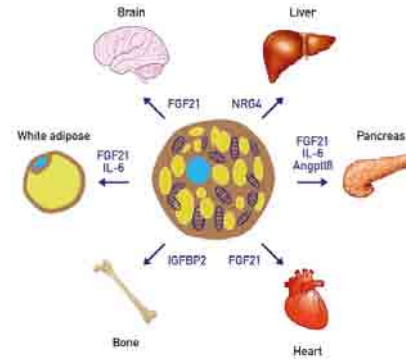
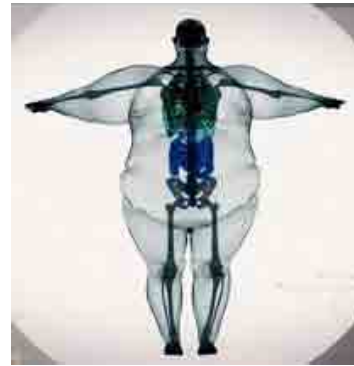
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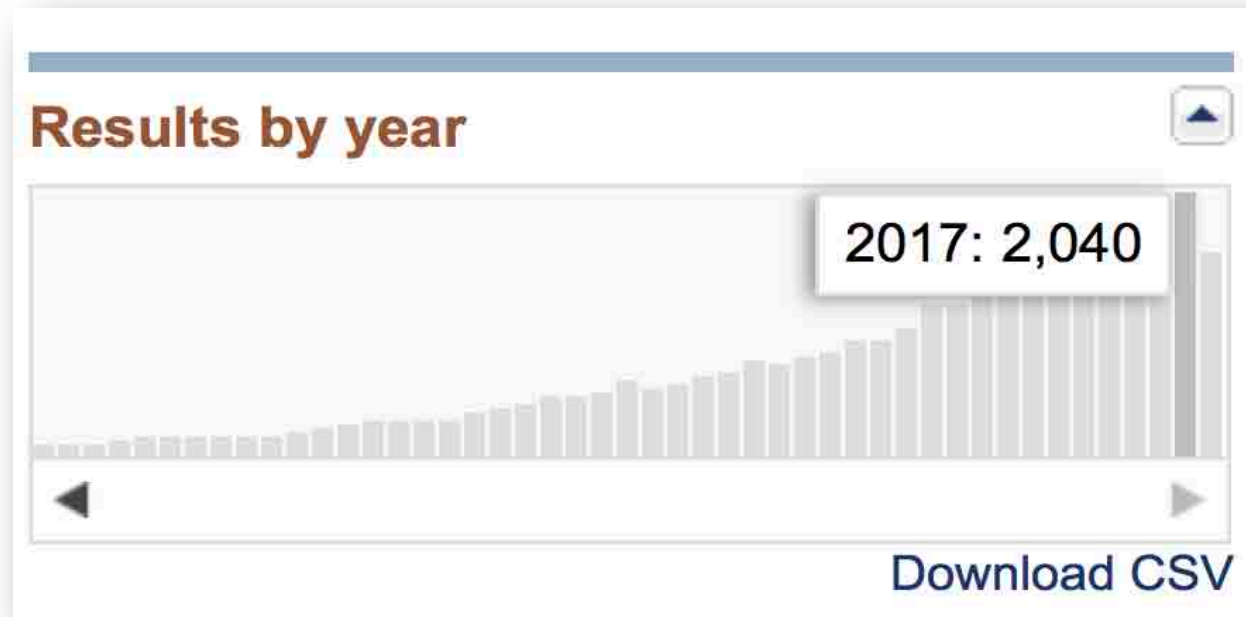
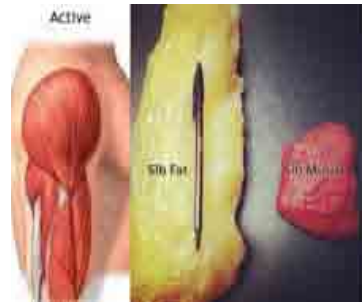
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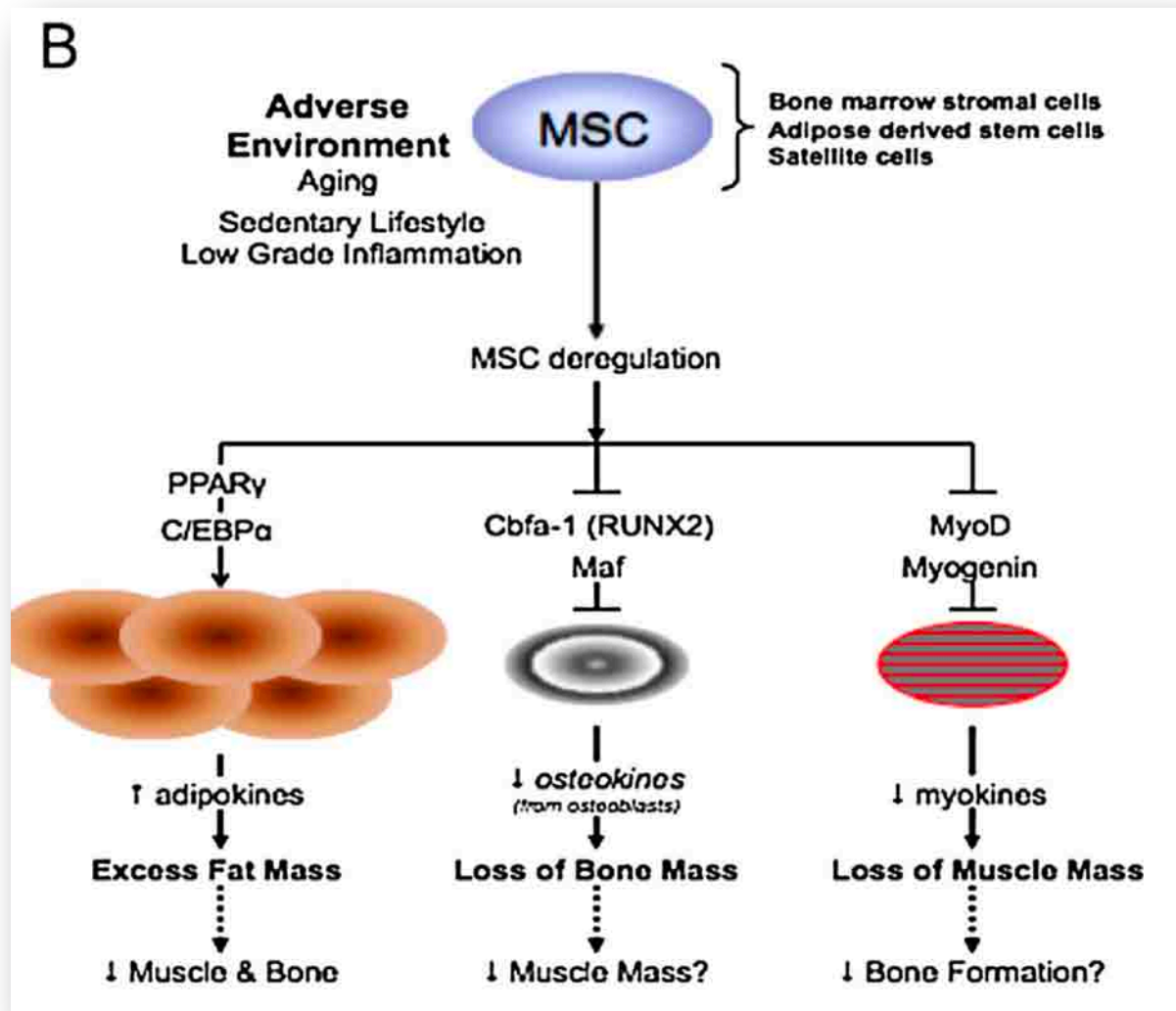


• Is the **Bone – Muscle – Fat Cross-talk** real ?

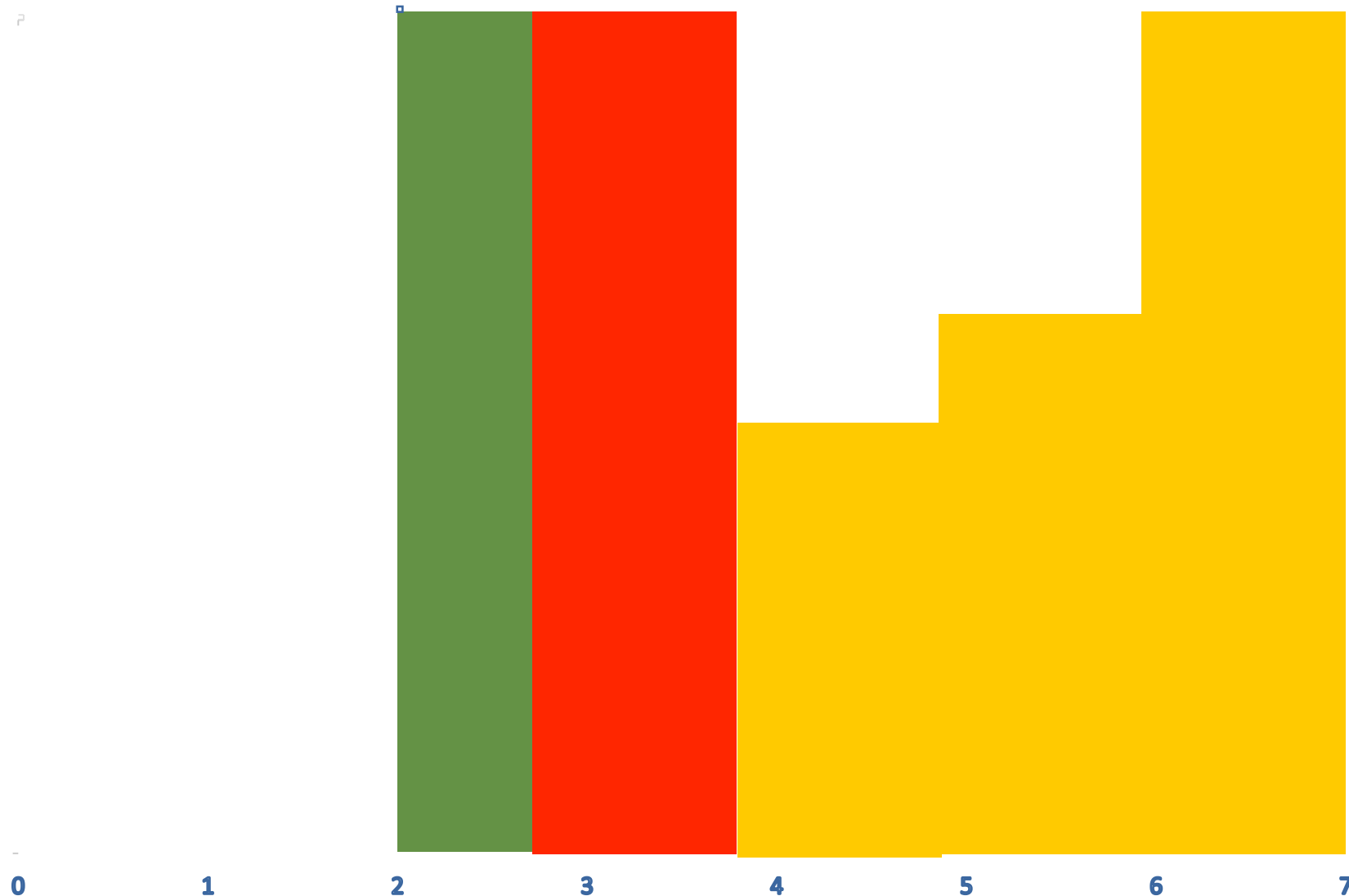




# TRANS-DIFFERENTIATION OF MSC TO DIFFERENT LINEAGES



# GROWTH PEAKS OF DIFFERENT TISSUES



**BONE:** peaks by 30 yrs, plateaus through young adulthood, and declines slowly with age. After menopause, 1-2% loss of bone mass/year.

**MUSCLE:** peaks at 30-40 yrs and gradually declines with age. At 70-80 yrs, subjects lose up 40% of mass.

**FAT:** fat tissue accumulation increases with aging and then it might plateau or decline in very old



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# BONE AND MUSCLE



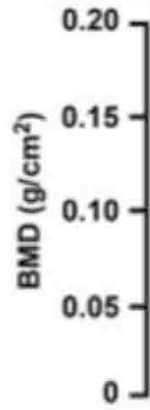


Table 3. Prevalence

Table 3. Prevalence of osteoporosis and osteopenia in a low-iron diet and alcohol consumption. \*P for trend.

Model 1	and alcohol
Normal	0.20
Osteopenia	0.15
Osteoporosis	0.10
Model 1	0.05
and alcohol	0

A



β-crosslaps (ng/ml)  
Total protein (g/dl)

### Reduction in proximal femoral strength due to long-duration spaceflight

J.H. Keyak<sup>a,b,c,\*</sup>, A.K. Koyama<sup>d</sup>, A. LeBlanc<sup>e,f</sup>, Y. Lu<sup>d</sup>, T.F. Lang<sup>d</sup>

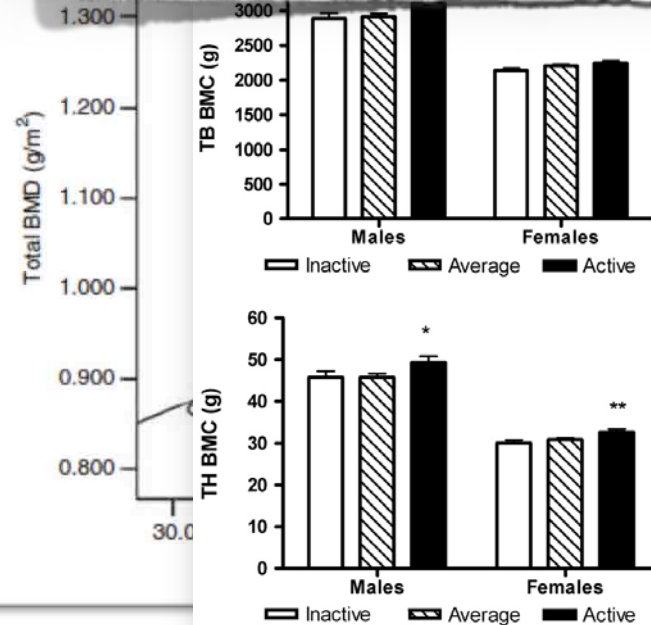
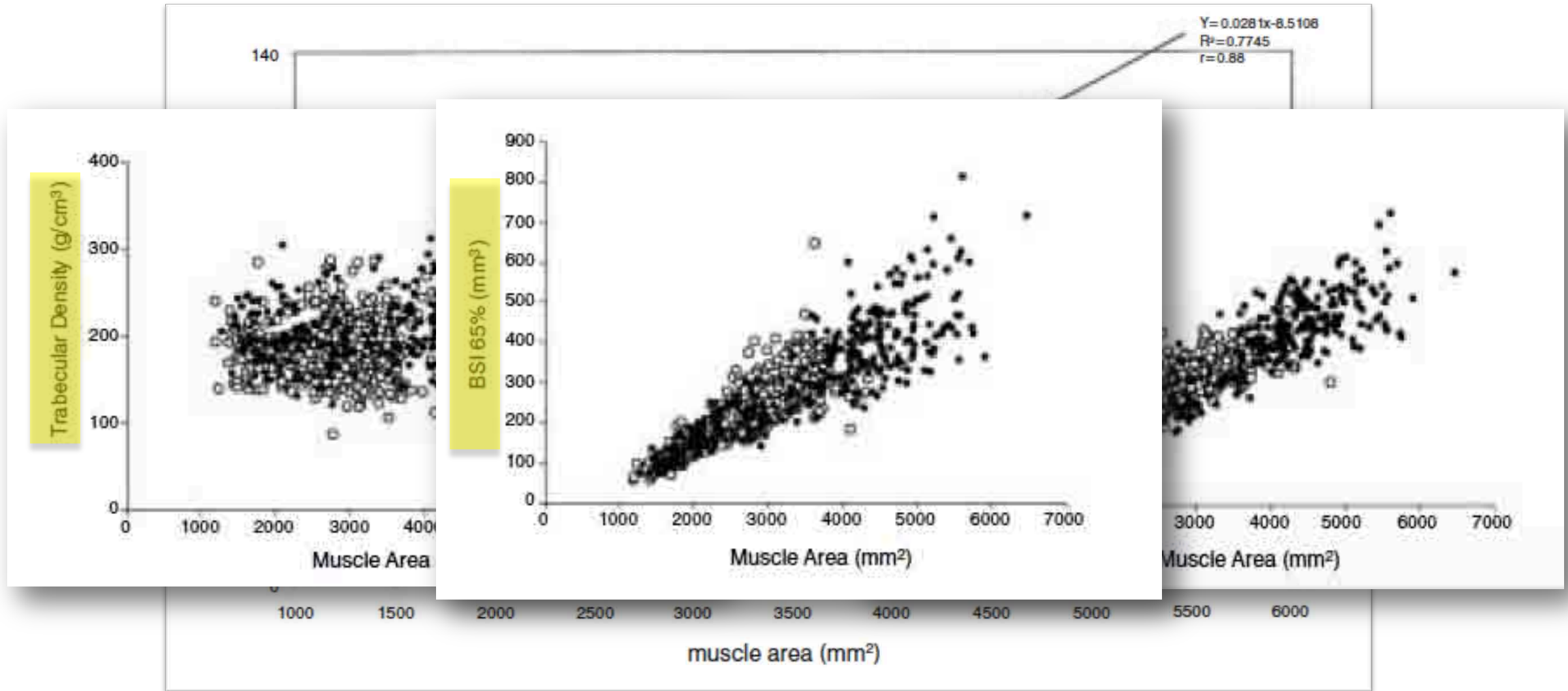


Table 2  
Rate of change in proximal femoral strength and DXA aBMD

Subject number	Change in F <sub>FE</sub> per month, stance loading	Change in F <sub>FE</sub> per month, fall loading	Change in DXA total femur aBMD per month [2]
1	-0.6%	0.6%	-0.7%
2	-2.1%	-3.3%	-0.9%
3	-1.4%	-2.7%	-1.0%
4	-5.0%	-0.7%	-1.6%
5	-2.2%	-2.3%	-1.1%
6	-4.0%	-1.8%	-1.0%
7	-1.4%	0.2%	-1.0%
8	-1.4%	-2.6%	-0.4%
9	-3.1%	-3.9%	-1.7%
10	-4.6%	-3.1%	-1.8%
11	-2.4%	-1.8%	-1.2%
12	-1.4%	-2.1%	-1.8%
13	-3.6%	-2.4%	-0.8%
Mean	-2.6%	-2.0%	-1.2%
Standard deviation	1.4%	1.3%	0.5%

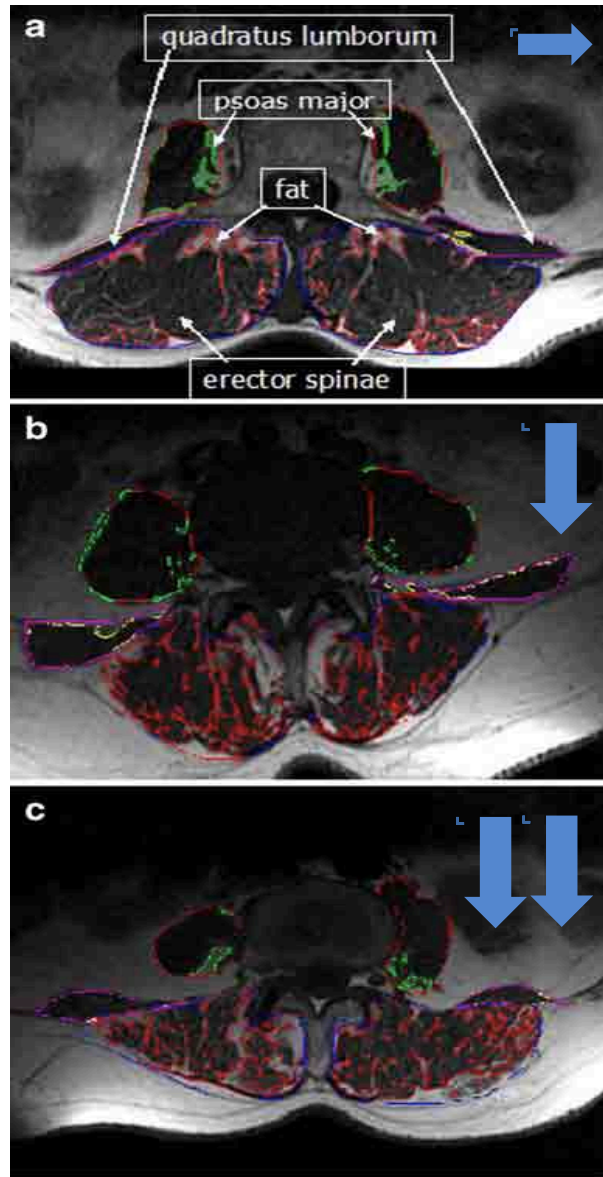


# MUSCLE SIZE/FORCE AND BMC/BONE STRENGTH





# DECLINE OF MUSCULAR STRENGTH AND BONE (1)



**Table 2** Correlation between lean muscle mass of paravertebral muscles, BMC, and age

Pearson correlation coefficient

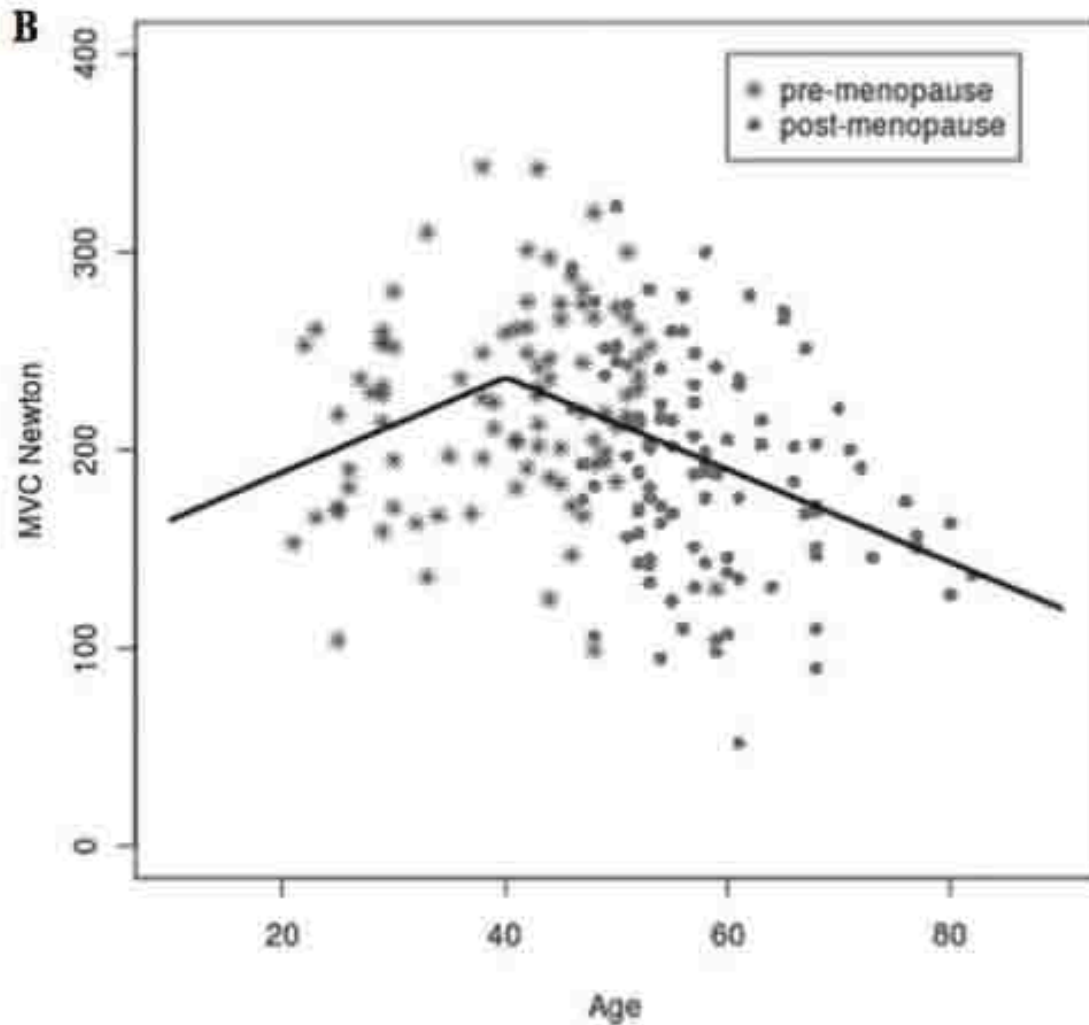
	Erector spinae	Psoas major	Qd. lumborum	Summated	Spine BMC
Spine BMC	0.28*	0.41*	0.36*	0.38*	1**
Age	-0.19*	-0.41**	-0.46**	-0.32**	-0.48**

As spine BMC decreases, muscle mass cross-sectional area decreases. As age increases, muscle mass decreases particularly in the psoas major and quadratus lumborum muscles

*Qd. lumborum* quadratus lumborum

\*  $p < 0.01$ , \*\*  $p < 0.001$

# DECLINE OF MUSCULAR STRENGTH AND BONE (2)



MVC: Maximal Voluntary Contraction ( Newton, N) by Hand Grip Dynamometer,

Table 2. Correlation matrix between MVC, densitometric and ultrasonometric parameters in the whole sample and in the sample subdivided according to gonadal status

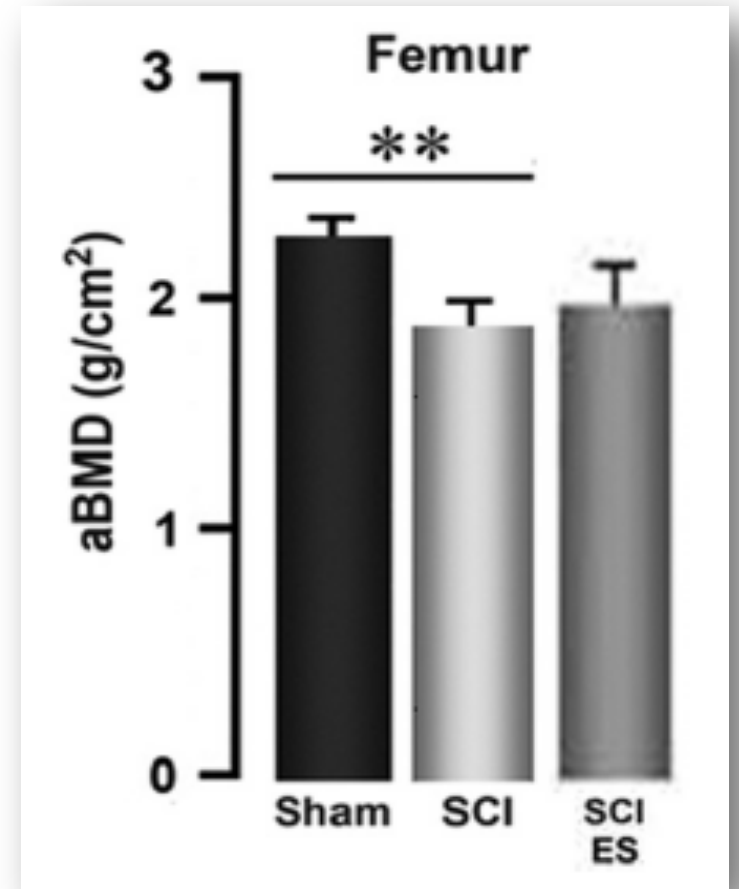
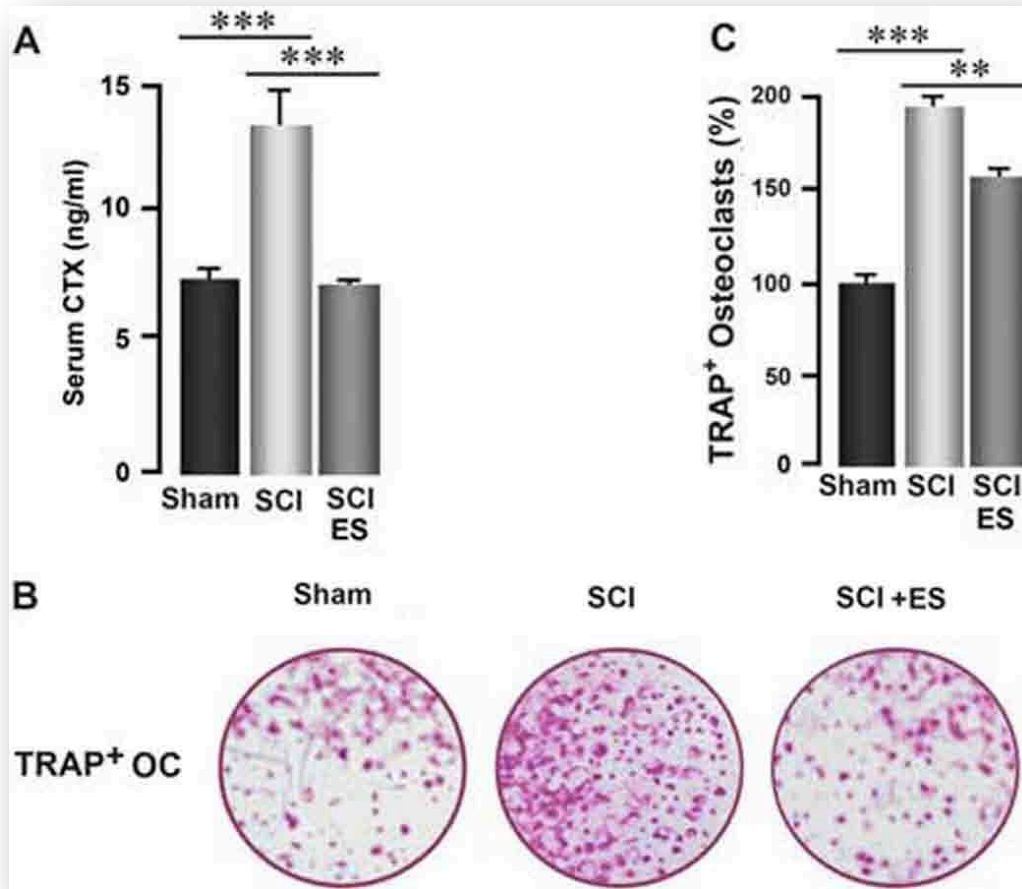
Parameters		Whole sample (n=194)	Premenopausal (n=92)	Postmenopausal (n=102)
		MVC <sup>a</sup> (N)	MVC (N)	MVC (N)
<b>R-BMD<sup>b</sup></b>	r	0.354	0.111	0.354
<b>(g/cm<sup>2</sup>)</b>	p <sup>c</sup>	0.0001	0.290	0.0001
<b>ADSoS<sup>d</sup></b>	r	0.294	-0.187	0.307
<b>(m/s)</b>	p	0.0001	0.07	0.01
<b>UBPI<sup>e</sup></b>	r	0.311	-0.033	0.319
	p	0.0001	0.753	0.01

<sup>a</sup>Maximal voluntary contraction, <sup>b</sup>Bone mineral density at one third of the radius, <sup>c</sup>Spearman correlation coefficient, <sup>d</sup>Amplitude-dependent speed of sound, <sup>e</sup>Ultrasound bone profile index.

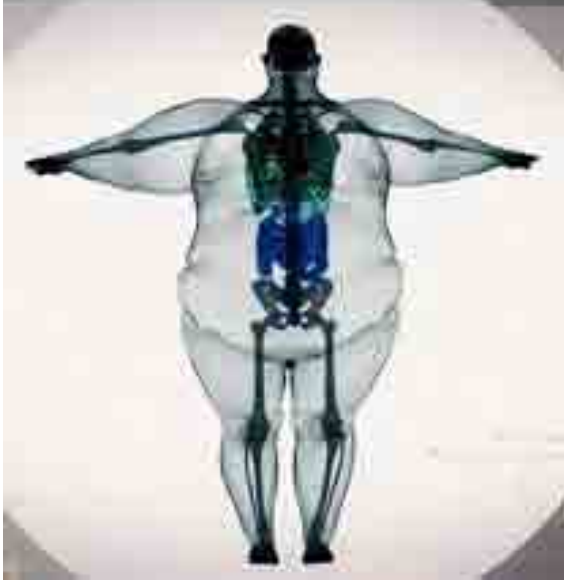
The decline of muscular strength is significantly correlated with quantitative and qualitative bone features



# ANTI-BONE-RESORPTIVE ACTIVITY OF MUSCLE CONTRACTION

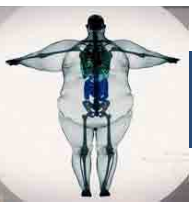


SCI: spinal cord injury; ES: electrical stimulation



# BONE AND FAT





# TRUNK/VISCERAL FAT AND BONE QUALITY



## Trabecular Bone Volume in Subjects from Each Tertile of Trunk Fat by DXA

Lowest Tertile



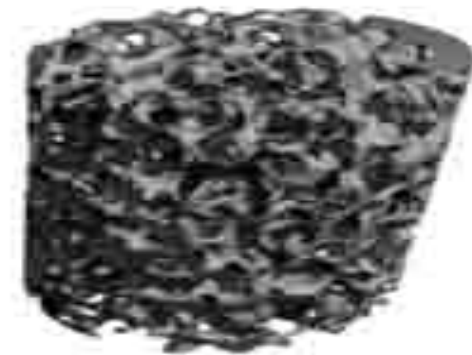
BMI: 22.2 kg/m<sup>2</sup>  
Trunk Fat: 16.8%  
LS Z score: -0.5  
BV/TV: 30.8%

Middle Tertile



BMI: 28.3 kg/m<sup>2</sup>  
Trunk Fat: 36.2%  
LS Z score: +1.4  
BV/TV: 22.5%

Highest Tertile

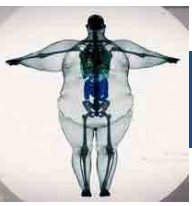


BMI: 36.9 kg/m<sup>2</sup>  
Trunk Fat: 42.0%  
LS Z score: +3.6  
BV/TV: 19.6%

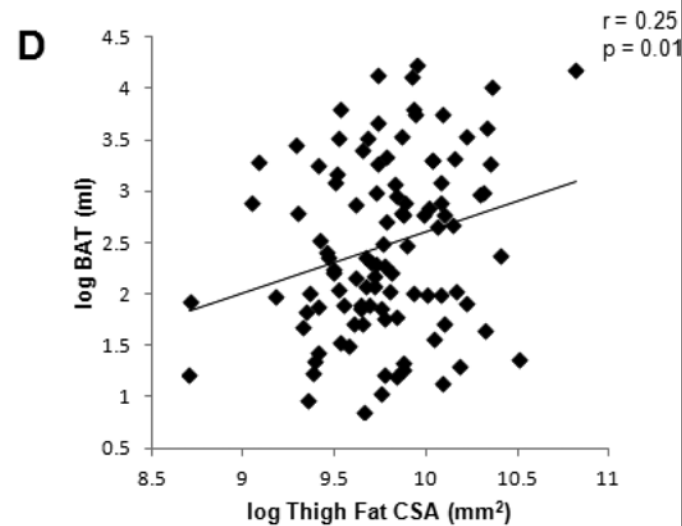
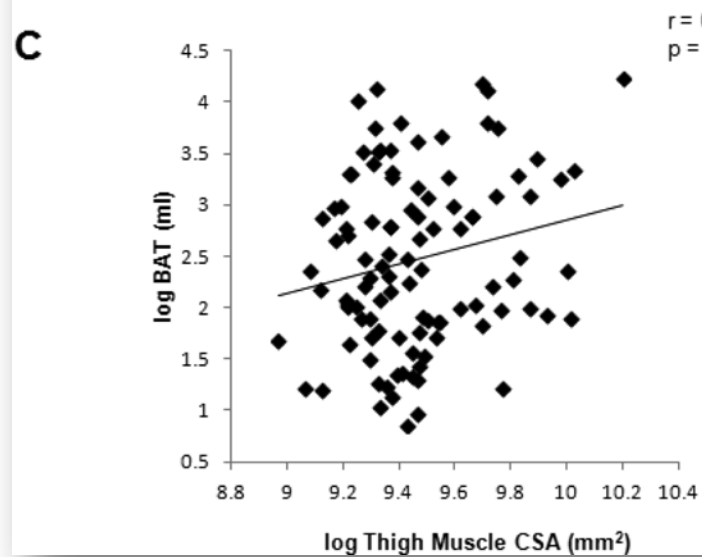
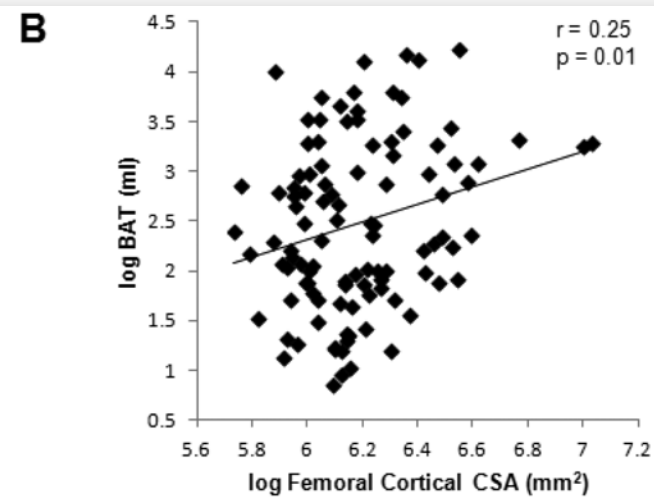
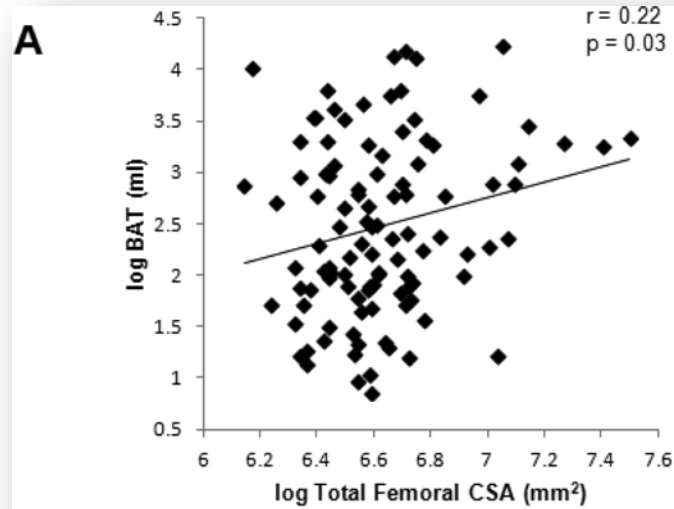
Trunk fat by DXA (%)

Trunk fat by DXA (%)

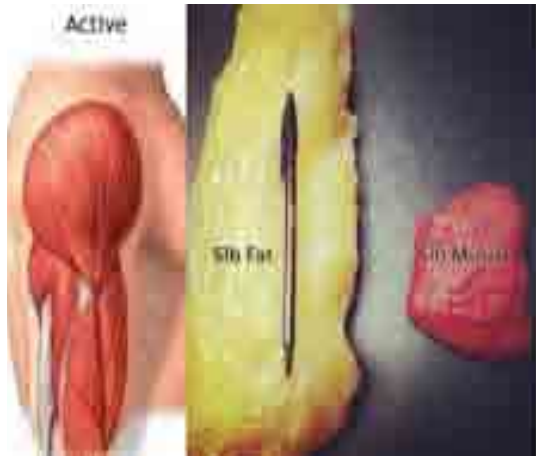
*Trabecular bone structure assessed by CT of transiliac crest bone biopsy samples obtained from representative subjects within each DXA trunk fat tertile. Subjects are chosen to represent the mean BV/TV seen in each tertile*



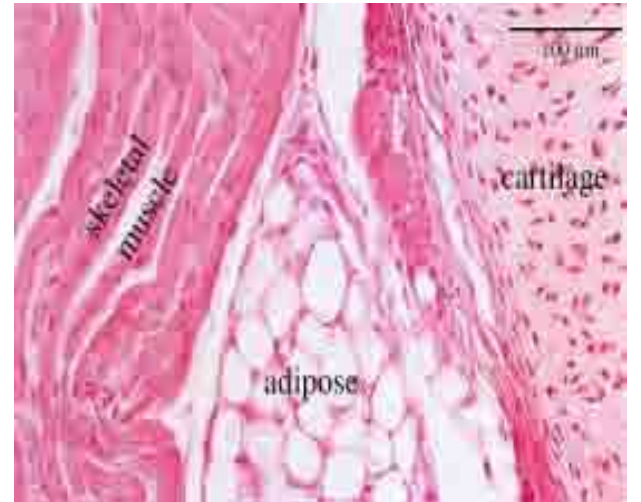
# BROWN ADIPOSE TISSUE and FEMORAL BONE STRUCTURE



**105 patients** (19 m, 86 f. mean age  $45 \pm 16$  years) who underwent 18-FDG PET/CT for benign etiologies ( $n=20$ ) or follow-up of successfully treated malignancies ( $n=85$ );



# MUSCLE AND FAT



# PHYSICAL ACTIVITY ON MUSCLE STRENGTH AND FAT INFILTRATION

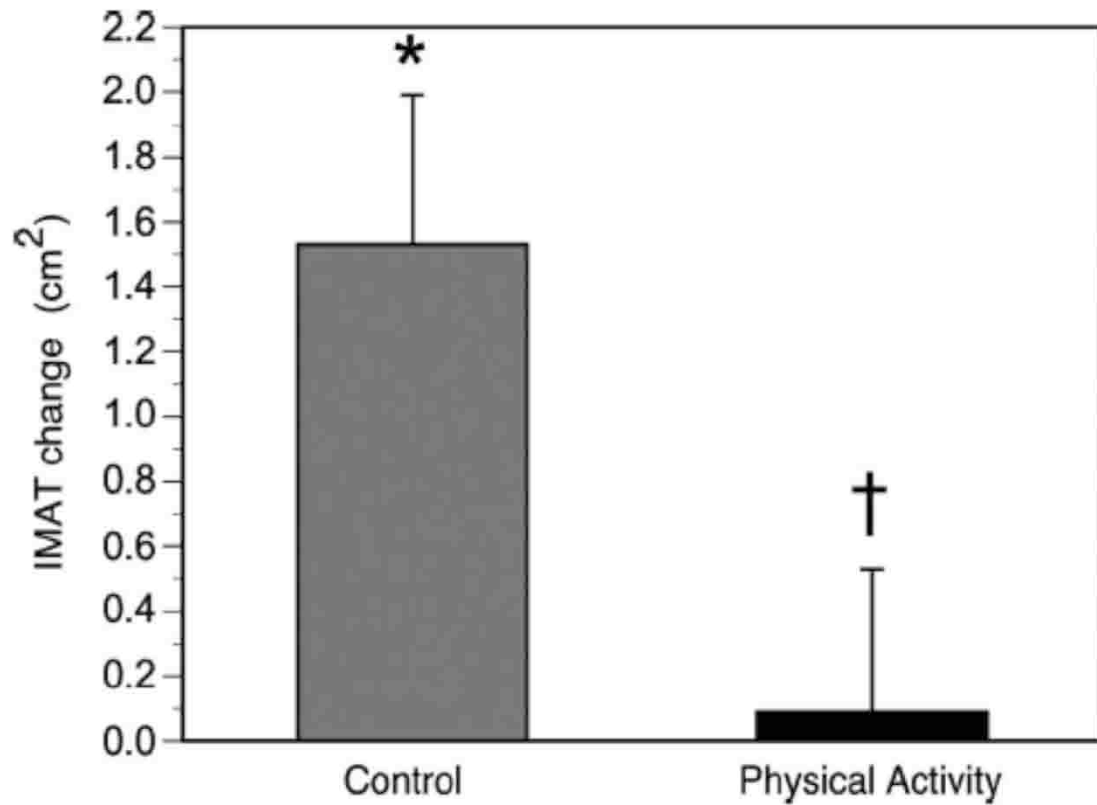
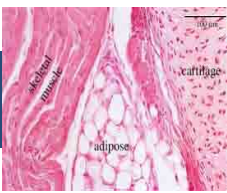
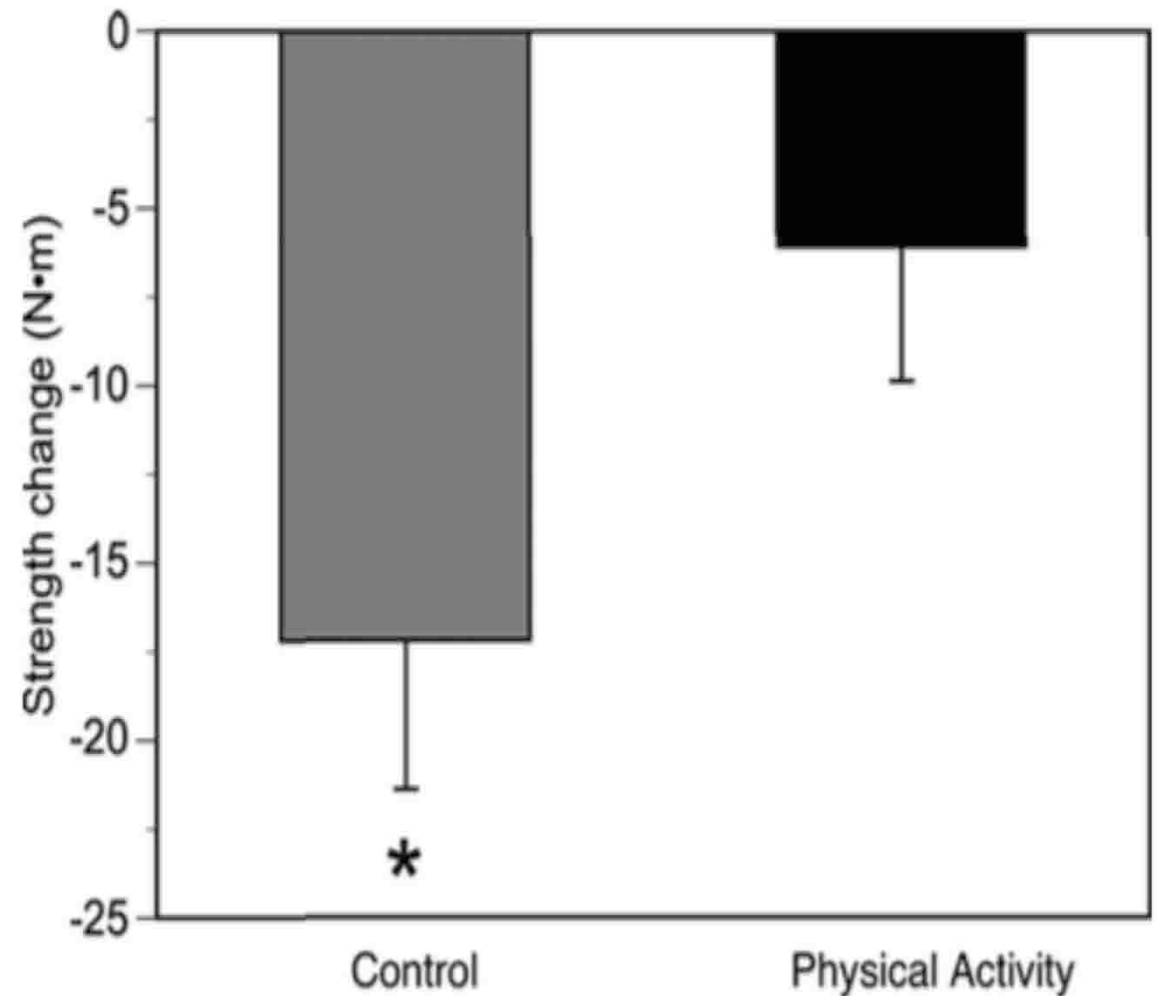


Fig. 2. Changes in mid thigh **intermuscular adipose tissue (IMAT)** in the physical activity and control groups. IMAT increased in the control group (within-group change: \* $P < 0.05$ , adjusting for multiple comparisons) but not in the physical activity group († $P < 0.05$  for between-group change).





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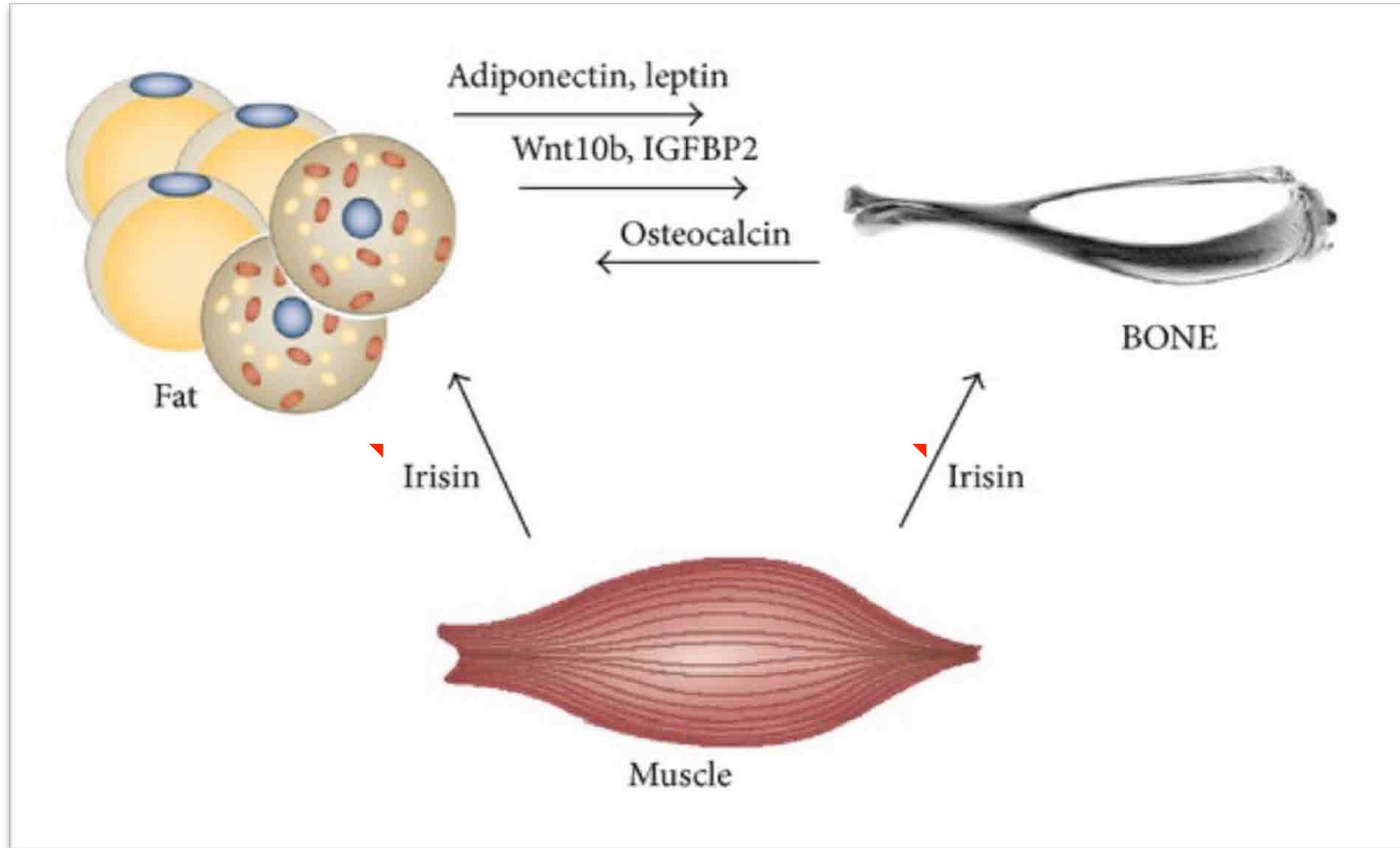
# MUSCLE – FAT – BONE AXIS

Factor	Brief description and effects on growth factors	Effect on muscle	Effect on fat	Effect on bone	Reference
PPAR $\gamma$	Required for adipogenesis, adipocyte maturation and metabolism, lipid accumulation, may cause trans-differentiation of pre-myocytes into adipocytes, retinoid X receptor (RXR) binding $\downarrow$ IGF-1, $\downarrow$ Runx2 (PPAR $\gamma$ $\downarrow$ Runx2), $\downarrow$ MyoD, $\downarrow$ myogenin	$\downarrow$ myogenesis	$\uparrow\uparrow$ adipogenesis	$\downarrow$ osteoblastogenesis	Ackert-Bicknell and Rosen, 2006; Aleshin et al., 2009; Hu et al., 1995; Lecka-Czernik and Suva, 2006; Metzger et al., 2005; Singh et al., 2007; Takazawa et al., 2009; Yu and Reddy, 2007
TNF $\alpha$	Regulator of cell metabolism, high levels produced by adipocytes (visceral), considered proinflammatory. Increases osteoclastogenesis (bone resorption) $\downarrow$ Runx2, $\downarrow$ osteocalcin, $\downarrow\downarrow$ MyoD, $\uparrow\uparrow$ myogenin, $\uparrow$ MAFbx, $\uparrow$ Murf-1	$\downarrow$ myogenesis	$\uparrow\uparrow$ adipogenesis	$\downarrow$ osteoblastogenesis	Cawthorn and Sethi, 2008; Chen et al., 2007; Hikiji et al., 2008; Langen et al., 2004; Mukai et al., 2007; Robinson et al., 2007; Zahorska-Markiewicz et al., 2000
Cbfa-1 (RUNX2)	Required for osteoblastogenesis, osteoblast maturation and differentiation, and genomic stability, enhanced by estrogen, regulated by PGE $_2$ $\uparrow\uparrow$ OC, ALP; $\uparrow$ aromatase, PPAR $\gamma$ ; $\downarrow$ Runx2	$\downarrow$ myogenesis	$\downarrow$ adipogenesis	$\uparrow\uparrow$ osteoblastogenesis	Isenmann et al., 2007; Jeong et al., 2010; McCarthy et al., 2003; Yu and Reddy, 2007; Zaidi et al., 2007
TGF $\beta$	Master cell regulatory factor, maintains MSC population, bone remodeling, elevated in obesity (visceral fat), regulated by PGE $_2$ $\uparrow$ Wnt pathway, $\downarrow$ PPAR $\gamma$	$\uparrow$ myogenesis	$\downarrow\downarrow$ adipogenesis	$\uparrow\uparrow$ osteoblastogenesis	Araujo-Jorge et al., 2012; Cao, 2007; Clarke and Liu, 2008; Fain, 2006; Ng et al., 2008; Zhao et al., 2008
IGF-1	Anabolic agent Reduced in obesity and osteoporosis $\uparrow\uparrow$ Myogenin, $\uparrow\uparrow$ MyoD	$\uparrow\uparrow$ myogenesis	$\downarrow$ adipogenesis	$\uparrow\uparrow$ osteoblastogenesis	Chen et al., 2007; Kaaks, 2004; Landin-Wilhelmsen et al., 1999; Sandhu et al., 2004
Maf	Osteoblast lineage commitment Aging and oxidative stress $\downarrow\downarrow$ Maf	$\downarrow$ myogenesis	$\downarrow\downarrow$ adipogenesis	$\uparrow\uparrow$ osteoblastogenesis	Nishikawa et al., 2010
MyoD	Required for myogenesis, differentiation of MSC/satellite cell to myocyte $\downarrow\downarrow$ TNF $\alpha$	$\uparrow\uparrow$ myogenesis	$\downarrow\downarrow$ adipogenesis	$\downarrow\downarrow$ osteoblastogenesis	Chen et al., 2007; Langen et al., 2004; Scheele et al., 2012
Myogenin	Required for myogenesis, differentiation of MSC/satellite cell to myocyte	$\uparrow\uparrow$ myogenesis	$\downarrow\downarrow$ adipogenesis	$\downarrow\downarrow$ osteoblastogenesis	Chen et al., 2007; Langen et al., 2004

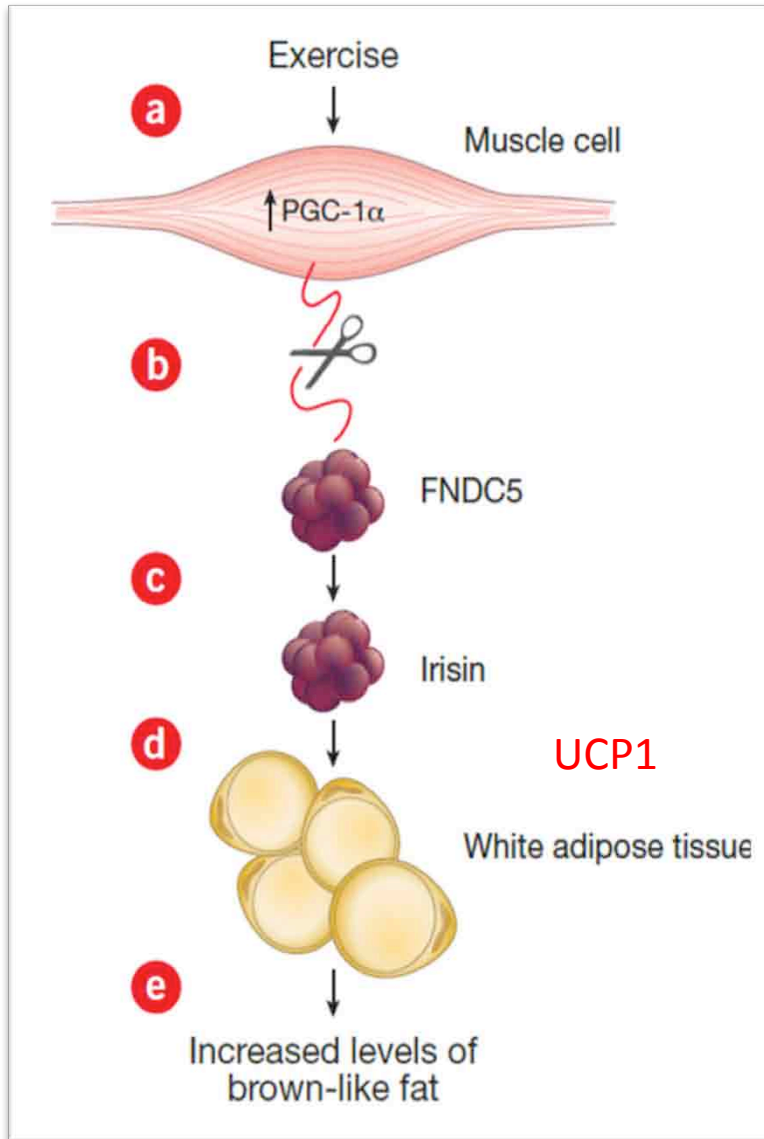
$\uparrow\uparrow$ : strong inducer,  $\downarrow\downarrow$ : strong inhibitor,  $\uparrow$ : inducer,  $\downarrow$ : inhibitor,  $\leftrightarrow$ : interdependent,  $\downarrow$ : inversely proportional



# MYOKINE: IRISIN



# MYOKINE: IRISIN



researchers have identified a new hormone, **irisin**, which mediates some of the benefits exercise has on metabolism.

Exercise induces increased expression of peroxisome proliferation-activated receptor- $\gamma$  coactivator 1 $\alpha$  (PPARGC1A; PGC-1 $\alpha$ ) [a], which boosts expression of the membrane protein **fibronectin type III domain containing 5 (FNDC5)** [b]. **FNDC5 is proteolytically cleaved, resulting in the release of irisin** [c], which is carried in the blood to **white adipose tissue** [d], where it stimulates the browning of white fat [e].

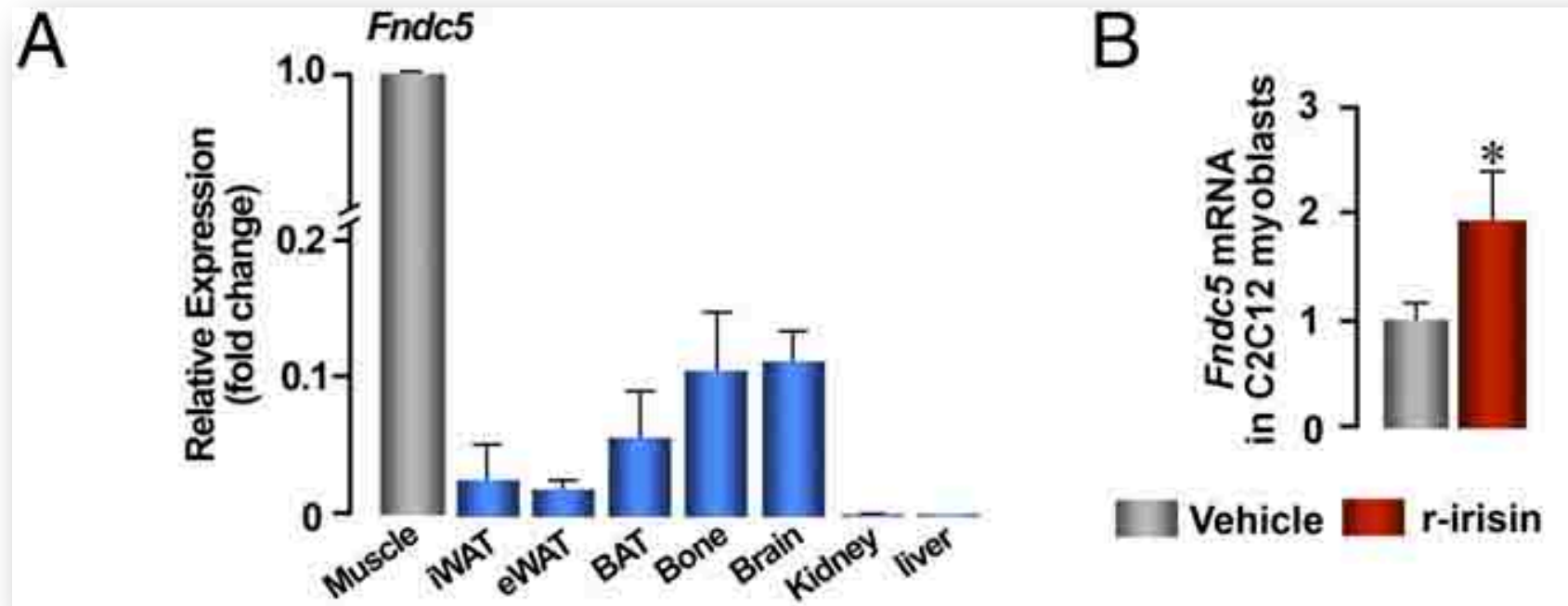
In mice fed a high-fat diet, greater expression of irisin decreased weight gain and increased glucose tolerance compared with normal irisin expression.

## The myokine irisin increases cortical bone mass

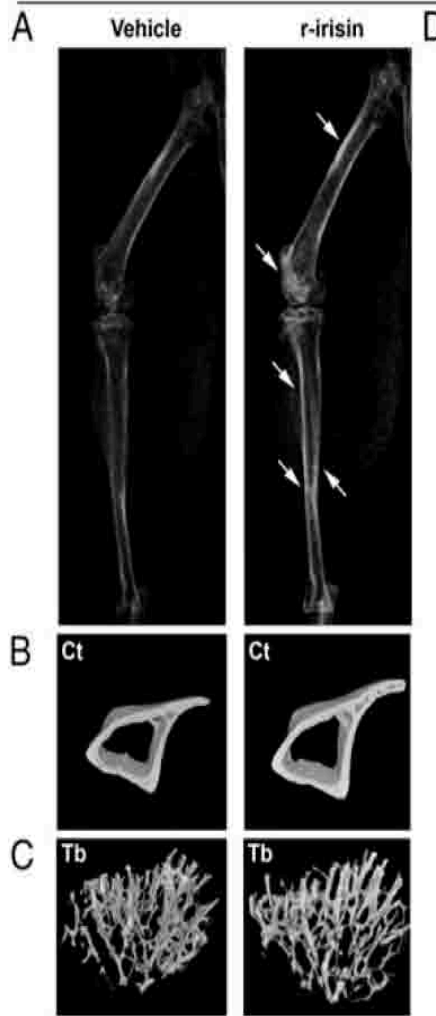
Graziana Colaianni<sup>h,1</sup>, Concetta Cuscito<sup>h,1</sup>, Teresa Mongelli<sup>a</sup>, Paolo Pignataro<sup>a</sup>, Cinzia Buccoliero<sup>a</sup>, Peng Liu<sup>b</sup>, Ping Lu<sup>b</sup>, Loris Sartini<sup>c</sup>, Mariasevera Di Comite<sup>a</sup>, Giorgio Mori<sup>d</sup>, Adriana Di Benedetto<sup>d</sup>, Giacomina Brunetti<sup>a</sup>, Tony Yuen<sup>b</sup>, Li Sun<sup>b</sup>, Janne E. Reseland<sup>e</sup>, Silvia Colucci<sup>a</sup>, Maria I. New<sup>b,2</sup>, Mone Zaidi<sup>b,2</sup>, Saverio Cinti<sup>a</sup>, and Maria Grano<sup>a,2</sup>

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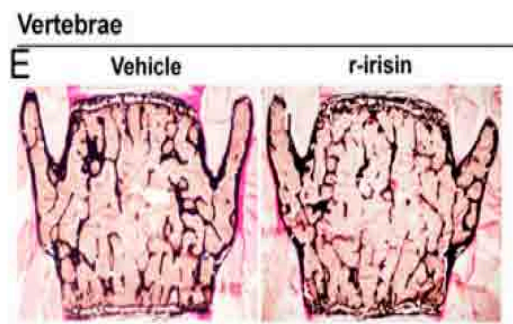
Contributed by Maria I. New, August 27, 2015 (sent for review May 21, 2015; reviewed by Christopher Huang and Carlos M. Isales)



Tibia

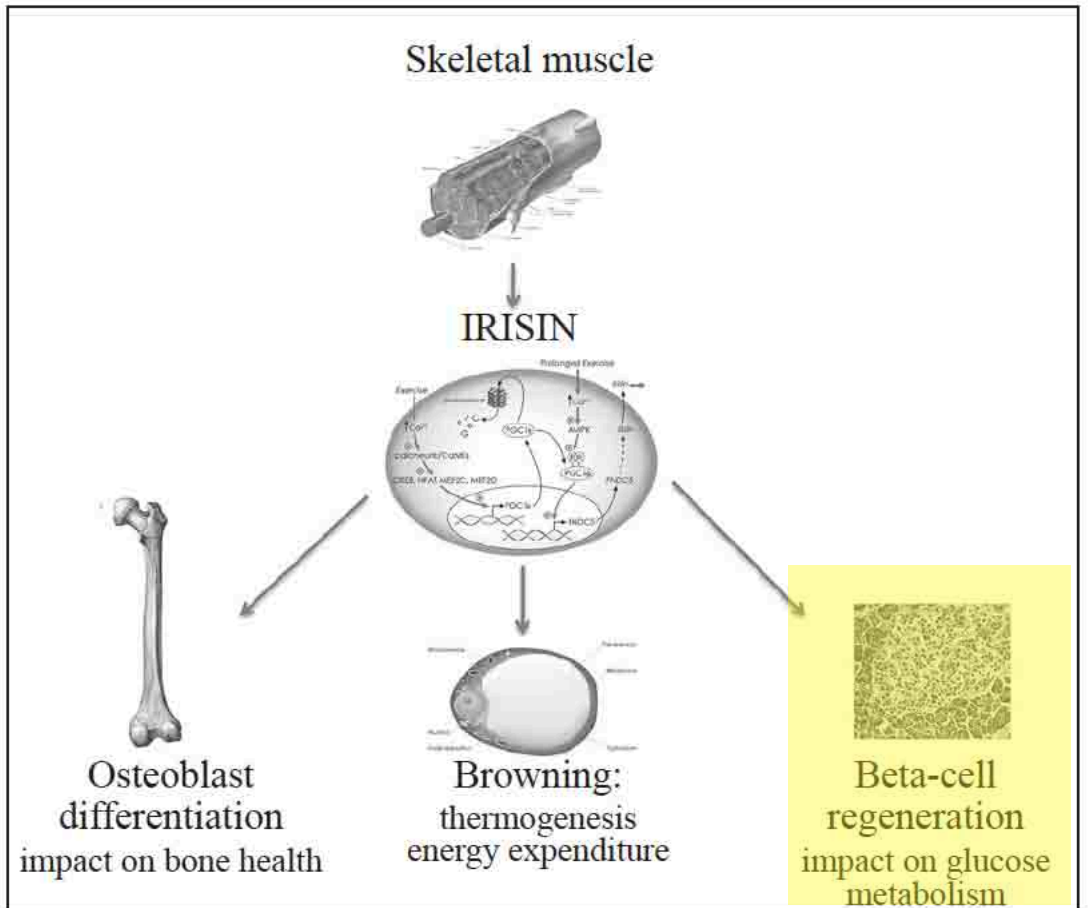


Injection	Vehicle	r-irisin	P value
TMD Cortical (gHA/cm <sup>3</sup> )	1.01 ±0.02	1.08 ±0.01	0.01
pMOI (mm <sup>4</sup> )	0.42 ±0.02	0.50 ±0.01	0.01
Ct.BS (mm <sup>2</sup> )	14.13 ±0.26	15.20 ±0.27	0.02
Ct.Pm (mm)	12.01 ±0.23	12.89 ±0.26	0.03
Tt.Area (mm <sup>2</sup> )	1.71 ±0.02	1.80 ±0.03	0.04
Marrow Area (mm <sup>2</sup> )	0.96 ±0.02	1.05 ±0.03	0.03
Ct.Th (mm)	0.13 ±0.004	0.12 ±0.01	0.62
BMD (gHA/cm <sup>3</sup> )	0.13 ±0.01	0.14 ±0.01	0.37
BV/TV (%)	51.13 ±2.26	44.32 ±2.60	0.07
Tb.Th (μm)	31.71 ±0.80	32.21 ±1.60	0.78
Tb.N (1/μm)	0.002 ±0.0001	0.001 ±0.0001	0.09
Tb.Sp (μm)	218.97 ±7.11	235.55 ±11.47	0.23



	Vehicle	r-irisin
BV/TV (%)	18.2 ± 1.15	17.6 ± 0.2
Tb.N (1/mm)	25.3 ± 0.7	26.1 ± 0.3
Tb.Th (mm)	0.7 ± 0.1	0.7 ± 0.0
Tb.Sp (mm)	3.3 ± 0.1	3.2 ± 0.0

# IRISIN AS A REGULATOR OF BONE AND GLUCOSE METABOLISM



Research Article

## Irisin Enhances Osteoblast Differentiation *In Vitro*

Graziana Colaianni,<sup>1</sup> Concetta Cuscito,<sup>1</sup> Teresa Mongelli,<sup>1</sup> Angela Oranger,<sup>1</sup>  
Giorgio Mori,<sup>2</sup> Giacomina Brunetti,<sup>1</sup> Silvia Colucci,<sup>1</sup> Saverio Cinti,<sup>3</sup> and Maria Grano<sup>1</sup>

Osteoporosis Int  
DOI 10.1007/s00198-014-2673-x

ORIGINAL ARTICLE

### Circulating irisin is associated with osteoporotic fractures in postmenopausal women with low bone mass but is not affected by either teriparatide or denosumab treatment for 3 months

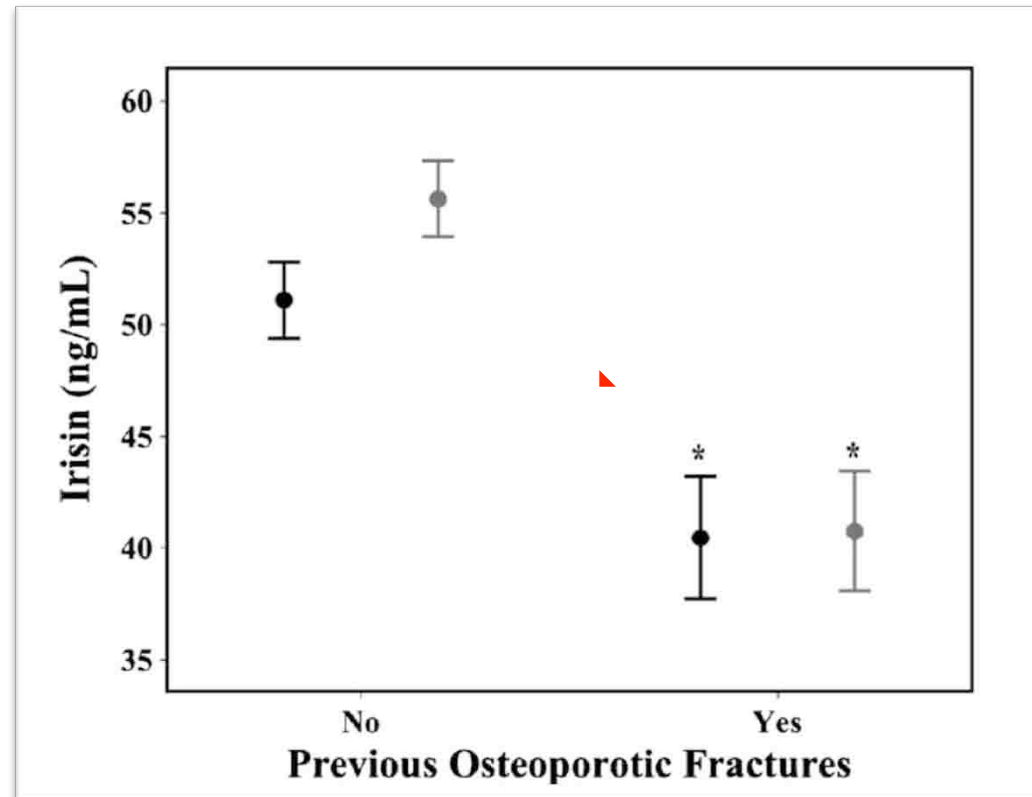
A. D. Anastasilakis · S. A. Polyzos · P. Makras ·  
A. Gkionisi · I. Bisbinas · A. Katsarou · A. Filippaios ·  
C. S. Mantzoros

### Irisin is associated with osteoporotic fractures independently of bone mineral density, body composition or daily physical activity

Andrea Palermo\*, Rocky Strollo\*, Ernesto Maddaloni\*, Dario Tuccinardi\*, Luca D'Onofrio\*,  
Silvia Irina Briganti\*, Giuseppe Defeudis\*, Mariangela De Pascalis\*, Maria Concetta Lazzaro\*,  
Georgia Colleluori\*, Silvia Manfrini\*, Paolo Pozzilli\*†‡ and Nicola Napoli Clinical Endocrinology (2015) 82, 615–619

## Circulating irisin is associated with osteoporotic fractures in postmenopausal women with low bone mass but is not affected by either teriparatide or denosumab treatment for 3 months

A. D. Anastasilakis · S. A. Polyzos · P. Makras ·  
A. Gkiomisi · I. Bishinas · A. Katsarou · A. Filippaios ·  
C. S. Mantzoros



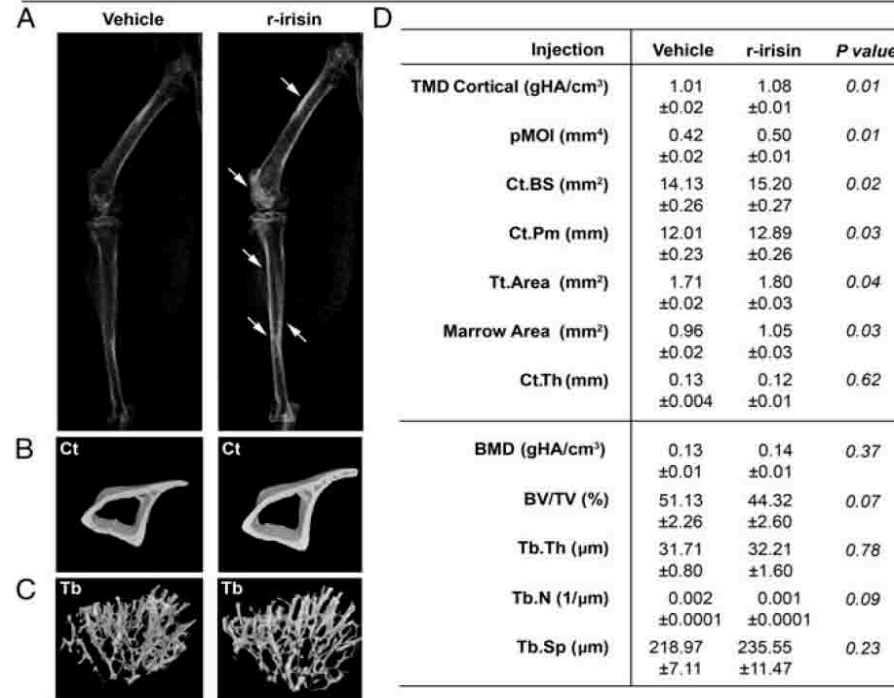
# The myokine irisin increases cortical bone mass

Graziana Colaianni<sup>a,1</sup>, Concetta Cuscito<sup>a,1</sup>, Teresa Mongelli<sup>a</sup>, Paolo Pignataro<sup>a</sup>, Cinzia Buccoliero<sup>a</sup>, Peng Liu<sup>b</sup>, Ping Lu<sup>b</sup>, Loris Sartini<sup>c</sup>, Mariasevera Di Comite<sup>a</sup>, Giorgio Mori<sup>d</sup>, Adriana Di Benedetto<sup>d</sup>, Giacomina Brunetti<sup>e</sup>, Tony Yuen<sup>b</sup>, Li Sun<sup>b</sup>, Janne E. Reseland<sup>a</sup>, Silvia Colucci<sup>b,2</sup>, Maria I. New<sup>b,2</sup>, Mone Zaidi<sup>b,2</sup>, Saverio Cinti<sup>a</sup>, and Maria Grano<sup>a,2</sup>

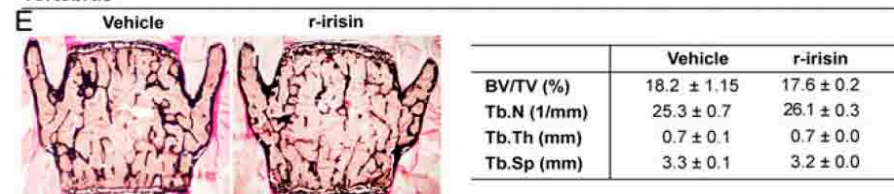
<sup>a</sup>Department of Basic Medical Science, Neuroscience and Sense Organs, University of Bari, 70124 Bari, Italy; <sup>b</sup>The Mount Sinai Bone Program, Departments of Medicine and Pediatrics, Mount Sinai School of Medicine, New York, NY 10029; <sup>c</sup>Department of Experimental and Clinical Medicine, Center of Obesity, United Hospitals, University of Ancona, 60020 Ancona, Italy; <sup>d</sup>Department of Clinical and Experimental Medicine, University of Foggia, 71100 Foggia, Italy; and <sup>e</sup>Department of Biomaterials, Institute for Clinical Dentistry, University of Oslo, Blindern, N-0317 Oslo, Norway

Contributed by Maria I. New, August 21, 2015 (sent for review May 21, 2015; reviewed by Christopher Huang and Carlos M. Isles)

## Tibia



## Vertebrae



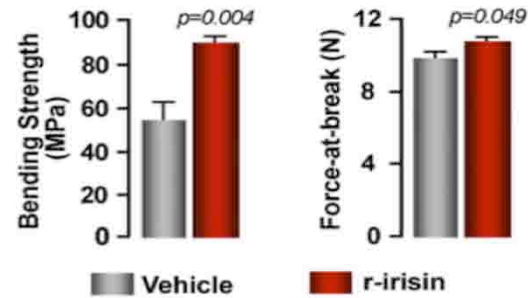
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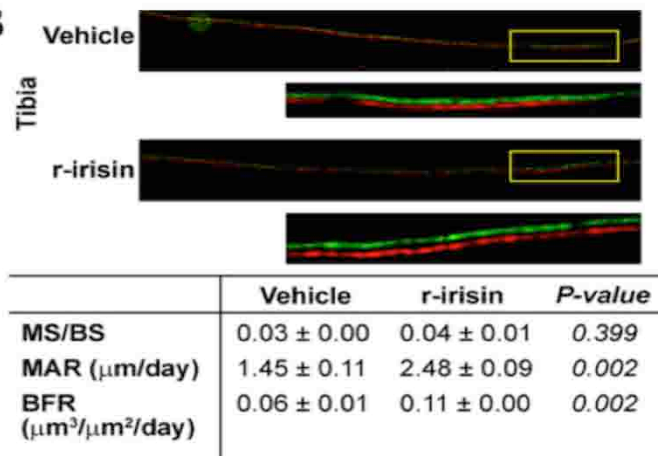
<sup>1</sup>Department of Basic Medical Science, Neuroscience and Sense Organs, University of Bari, 70124 Bari, Italy; <sup>2</sup>The Mount Sinai Bone Program, Departments of Medicine and Pediatrics, Mount Sinai School of Medicine, New York, NY 10029; <sup>3</sup>Department of Experimental and Clinical Medicine, Center of Obesity, United Hospitals, University of Ancona, 60020 Ancona, Italy; <sup>4</sup>Department of Clinical and Experimental Medicine, University of Foggia, 71100 Foggia, Italy; and <sup>5</sup>Department of Biomaterials, Institute for Clinical Dentistry, University of Oslo, Blindern, N-0317 Oslo, Norway

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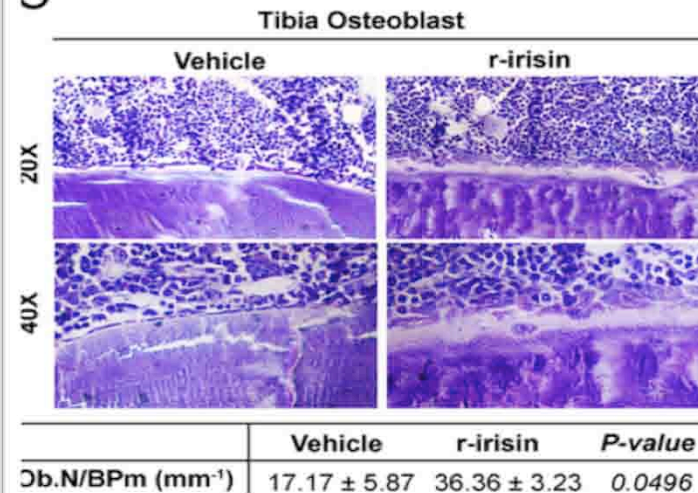
**A**



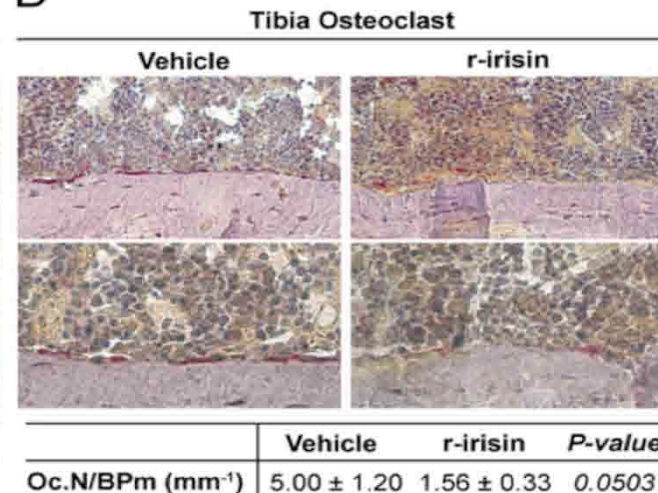
**B**



**C**



**D**



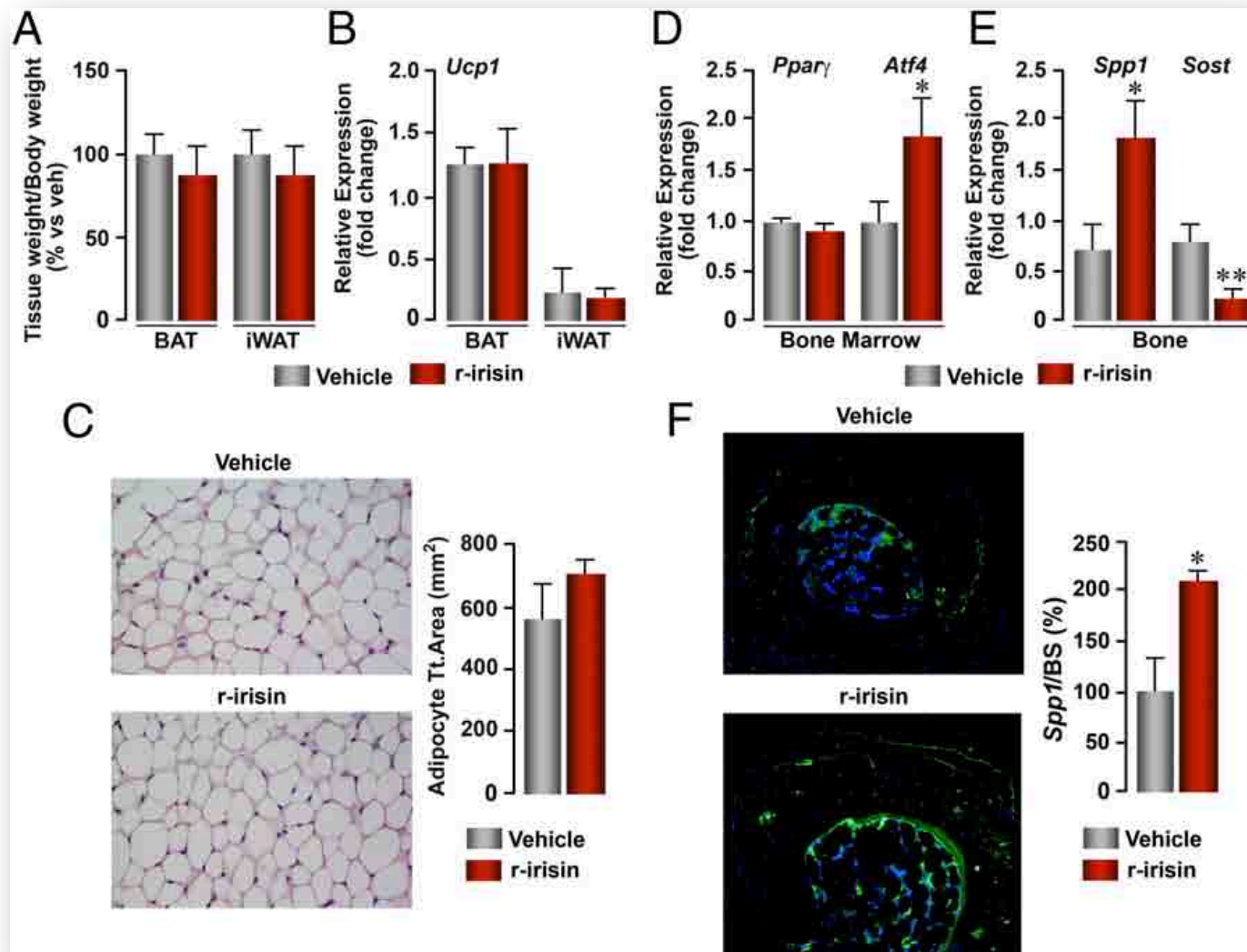


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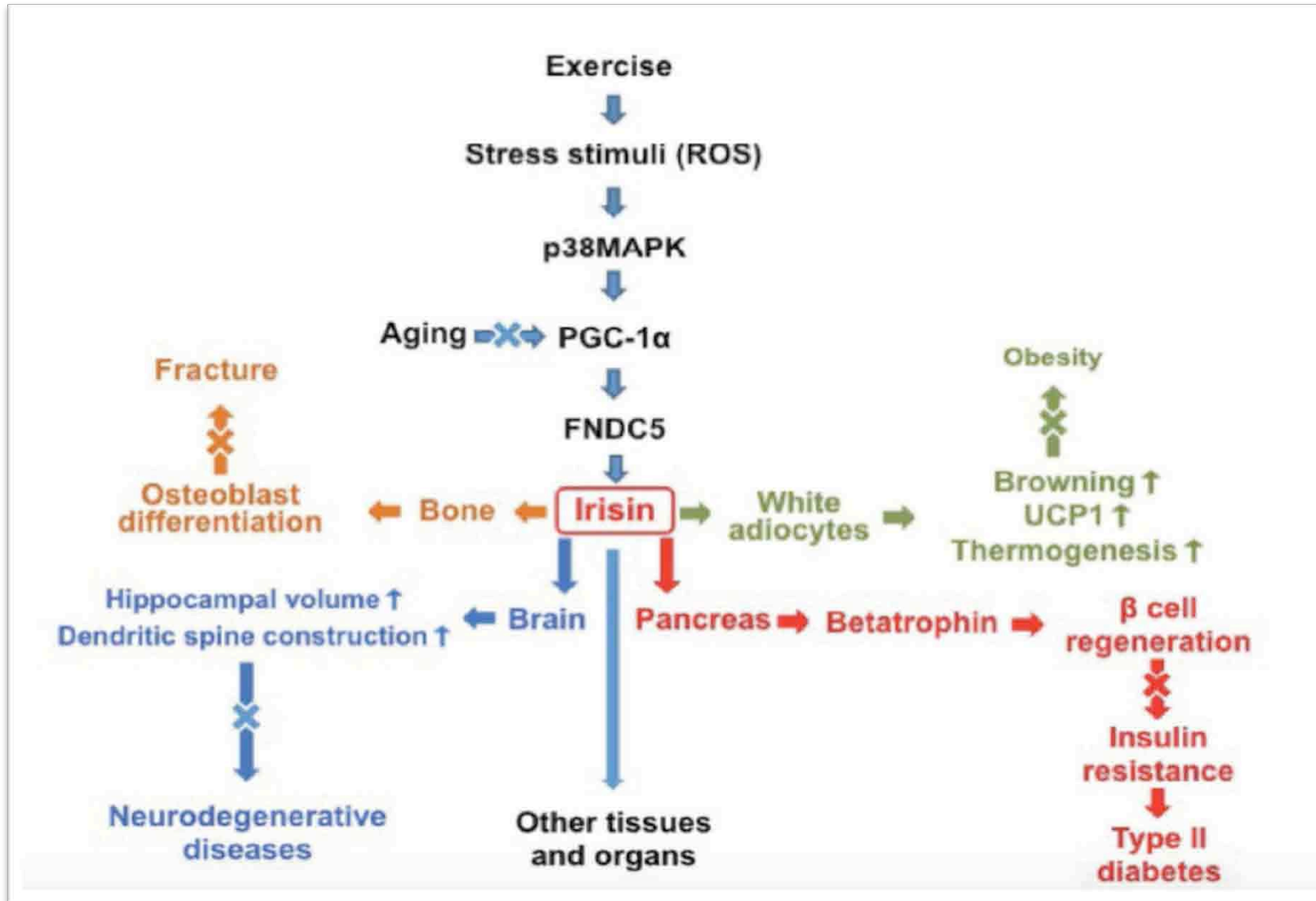
Graziana Colaianni<sup>1\*</sup>, Concetta Cuscito<sup>1\*</sup>, Teresa Mongelli<sup>1\*</sup>, Paolo Pignataro<sup>1\*</sup>, Cinzia Buccoliero<sup>1\*</sup>, Peng Liu<sup>2</sup>, Ping Lu<sup>2</sup>, Loris Sartini<sup>3</sup>, Mariasevera Di Comite<sup>4</sup>, Giorgio Mori<sup>5</sup>, Adriana Di Benedetto<sup>6</sup>, Giacomina Brunetti<sup>6</sup>, Tony Yuen<sup>2</sup>, Li Sun<sup>2</sup>, Janne E. Reseland<sup>7</sup>, Silvia Colucci<sup>1</sup>, Maria I. New<sup>1,2</sup>, Mone Zaidi<sup>1,2</sup>, Saverio Cinti<sup>1</sup>, and Maria Grano<sup>1,2</sup>

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# IRISIN: DIFFERENT METABOLIC ACTIONS



## **PTH excess may promote weight gain by impeding catecholamine-induced lipolysis-implications for the impact of calcium, vitamin D, and alcohol on body weight**

**Summary** Increased free intracellular calcium ( $[Ca^{2+}]_i$ ) in adipocytes blunts the lipolytic response to catecholamines by activating phosphodiesterase 3B – the same enzyme that mediates the antilipolytic effect of insulin – while also compromising the efficiency of insulin-stimulated glucose uptake. Physiological increases in parathyroid hormone (PTH) have been shown to increase  $[Ca^{2+}]_i$  in adipocytes. These considerations may rationalize recent evidence that high dietary intakes of calcium and/or dairy products may reduce risk for obesity, diabetes, and insulin-resistance syndrome, and they predict that other dietary measures which down-regulate PTH – such as good vitamin D status, and moderation in phosphate and salt intakes – may likewise be beneficial in these respects. Consistent with this position are reports that body weight is elevated in elderly subjects with both primary and secondary hyperparathyroidism; furthermore, insulin resistance is a well-known complication of both forms of hyperparathyroidism. The fact that regular alcohol consumption is associated with decreased PTH secretion may help to explain why moderate drinkers are less prone to insulin resistance, diabetes, and – in women – obesity. Down-regulation of PTH cannot be expected to promote dramatic weight loss, but in the long-term it may lessen risk for significant weight gain and diabetes, and conceivably may potentiate the fat loss achievable with caloric restriction and/or exercise.

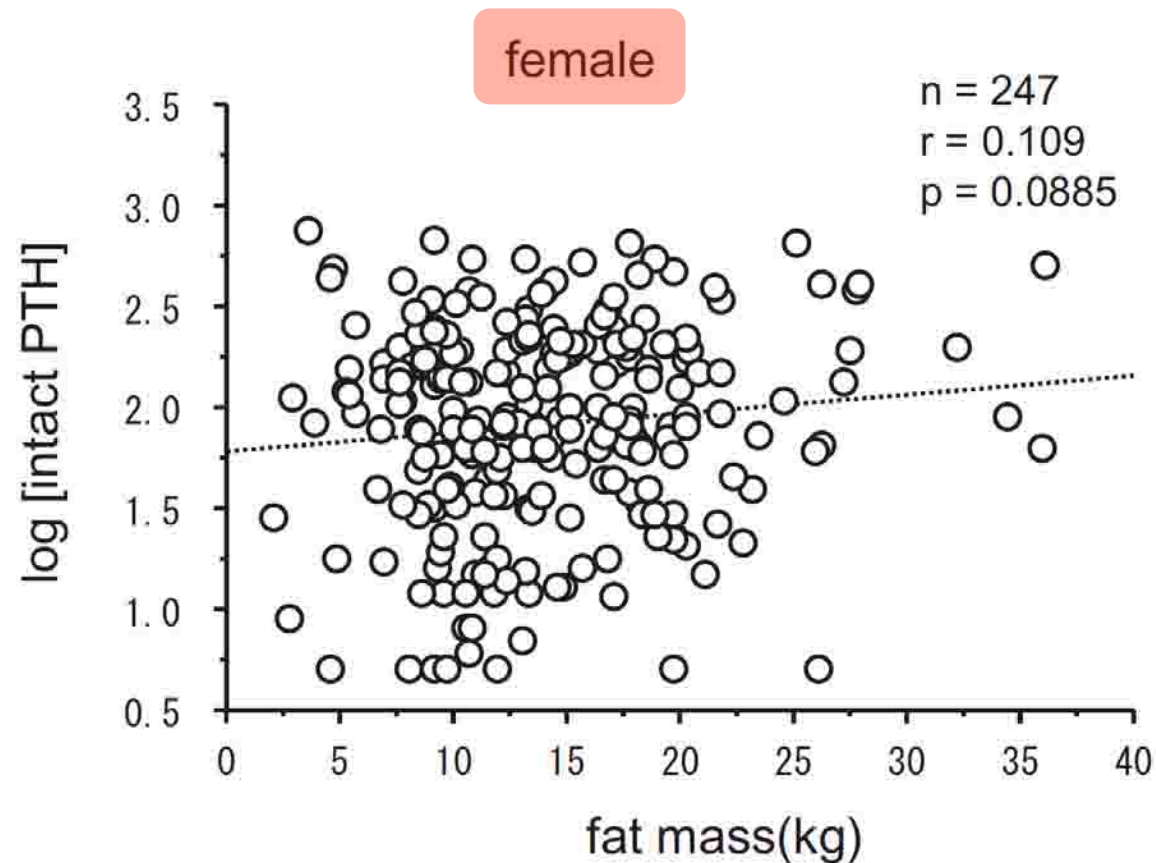
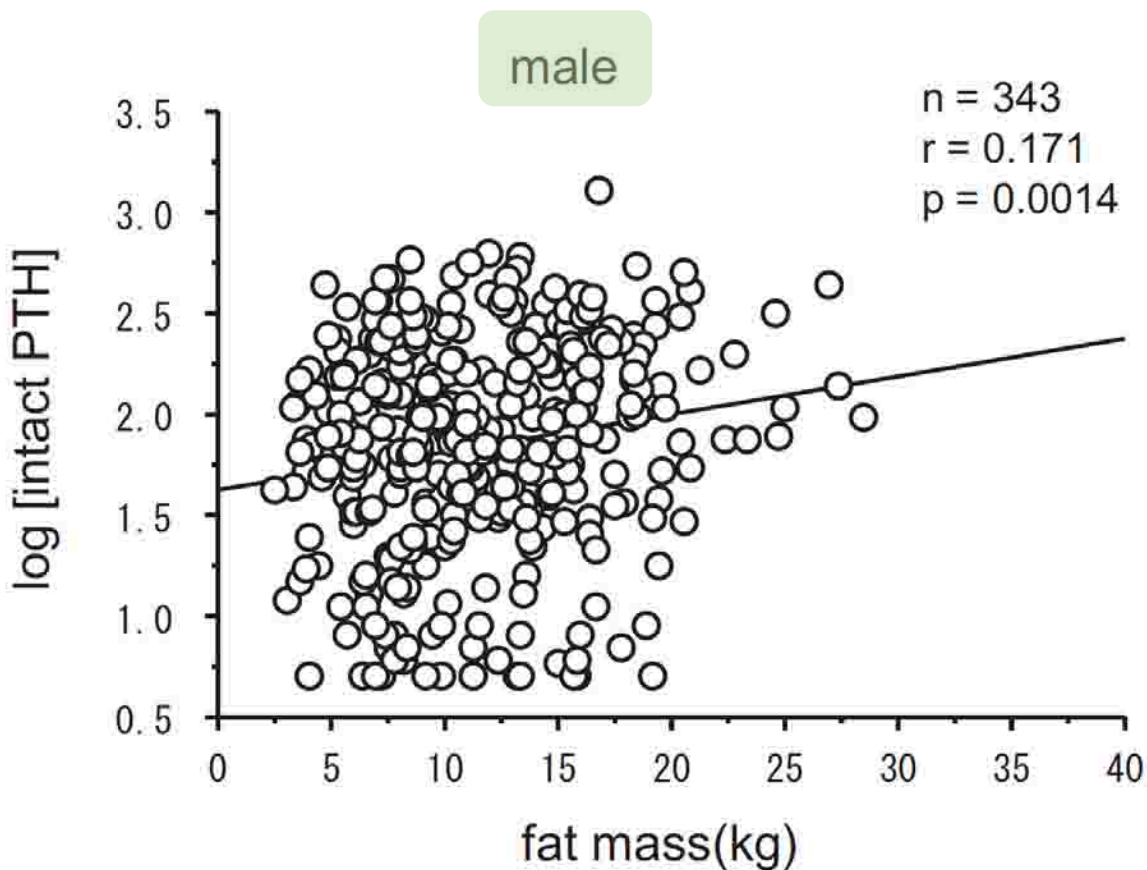
## Parathyroid hormone is significantly associated with body fat compartment in men but not in women following a hip fracture

**Table 3** Linear regression analysis in the 57 men and in the 518 women

Independent variables	$\beta$	Partial correlation	$p$
<b>Analysis in the 57 men</b> ( $r^2 = 0.43$ ; $F = 13.53$ ; $p < 0.001$ )			
PTH	<b>0.46</b>	<b>0.51</b>	<b>&lt;0.001</b>
Age	<b>-0.33</b>	<b>-0.38</b>	<b>0.004</b>
Phosphate	0.15	0.19	0.16
Albumin-adjusted calcium	-0.02	-0.03	0.85
Estimated GFR	-0.02	-0.03	0.83
Albumin	<b>-0.22</b>	<b>-0.27</b>	<b>0.047</b>
Barthel Index score	0.12	0.14	0.31

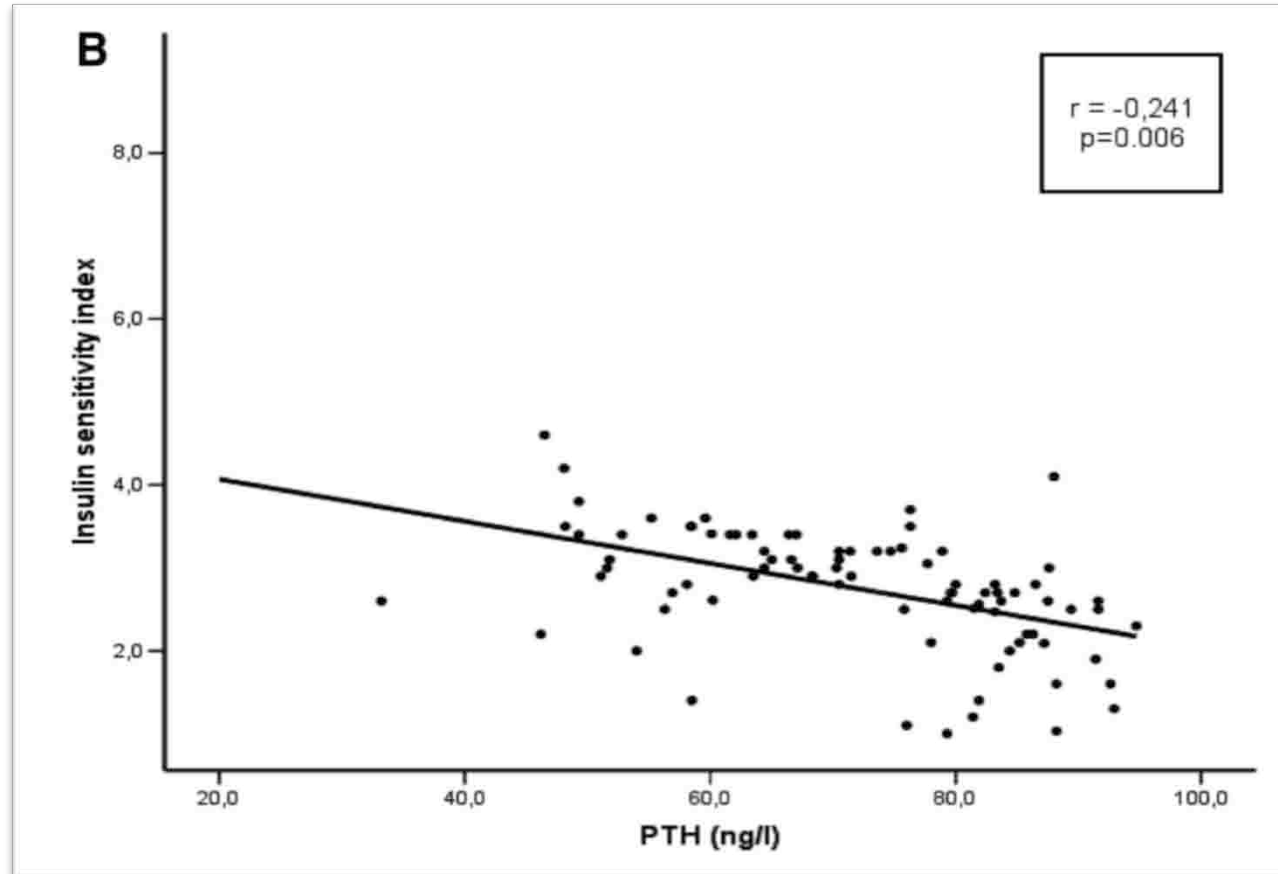
**Results** In the 57 men, we found a significant correlation between PTH and both body mass index (BMI) ( $\rho = 0.37$ ;  $p = 0.020$ ) and trunk fat percentage ( $\rho = 0.62$ ;  $p < 0.001$ ). After multiple adjustments, we confirmed a significant association between PTH and BMI ( $r = 0.38$ ;  $p = 0.004$ ) or trunk fat percentage ( $r = 0.51$ ;  $p < 0.001$ ). In the 518 women, we found a slightly significant correlation between PTH and BMI ( $\rho = 0.09$ ;  $p = 0.047$ ), but after adjustments the correlation coefficient dropped to

## Significant Positive Association Between Parathyroid Hormone and Fat Mass and Lean Mass in Chronic Hemodialysis Patients

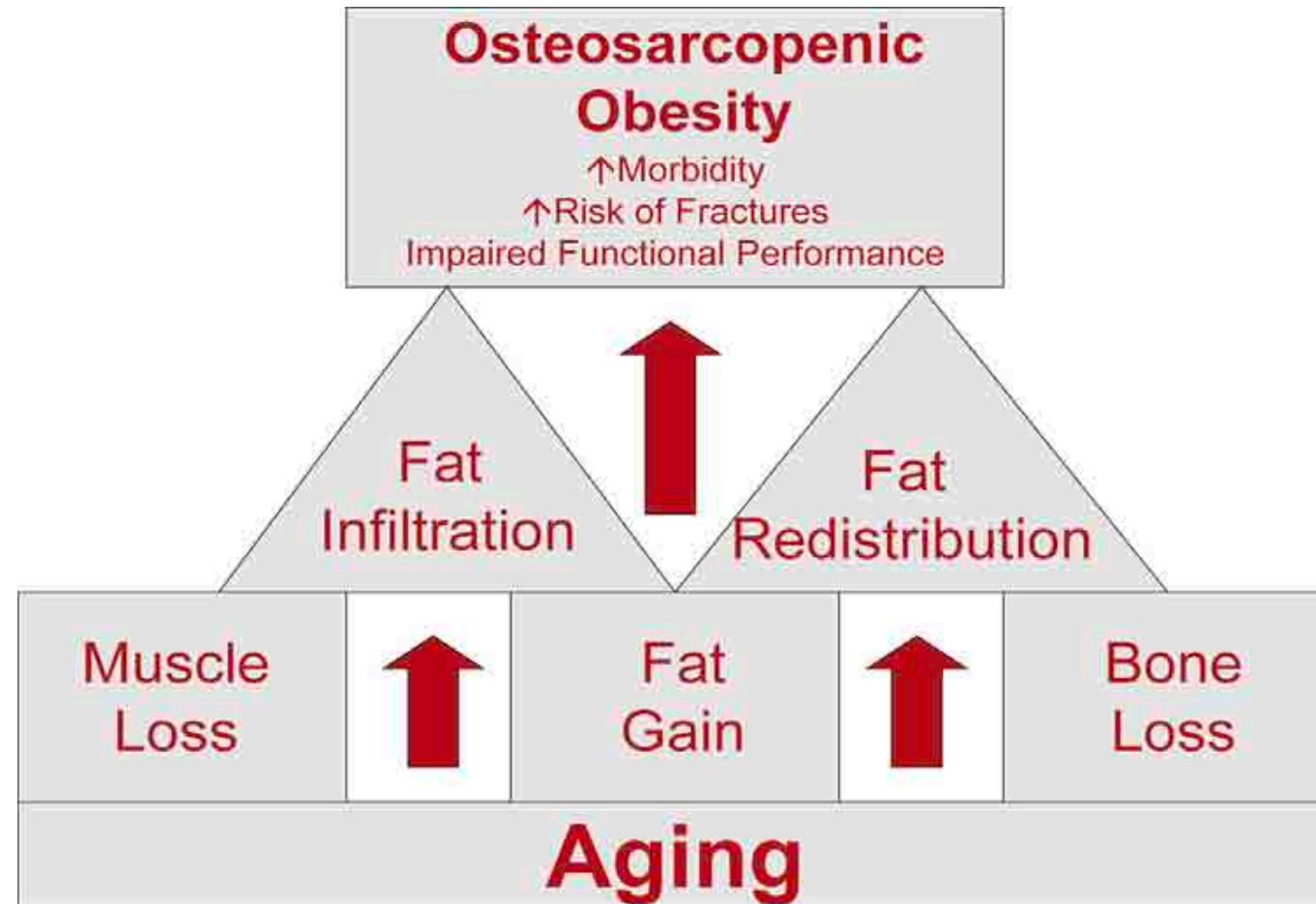


# Parathyroid Hormone and Insulin Resistance in Distinct Phenotypes of Severe Obesity: A Cross-Sectional Analysis in Middle-Aged Men and Premenopausal Women

J Clin Endocrinol Metab, December 2012, 97(12):4724–4732



# TAKE HOME MESSAGES



THANK YOU

